

PROCEDURE FOR FECAL EXAMINATION BY CENTRIFUGATION

1. Mix 2—3 g of fresh feces with 15 ml of the following sugar solution in a disposable cup until there is uniform consistency. Sugar solution recipe: Mix 1 lb. sugar with 350 ml of boiling water until the sugar has dissolved (specific gravity, approximately 1.25-1.27). (6.6 ml of phenol may be added as a preservative.)
2. Strain through cheesecloth into a conical 15-ml centrifuge tube. Fill the tube even with the top or with a slight reverse meniscus.
3. Carefully place the tube into a "swinging-bucket" centrifuge, using a proper balance tube opposite it.
4. Apply a coverslip to the top of the tube and gently "seat" with finger pressure.
5. Centrifuge at 1,200-1,500 rpm for 5-10 minutes with coverslip in place.
6. Lift coverslip directly upward and immediately place on microscope slide.
7. Scan the entire slide for parasites under 10X objective and for protozoa under 40x.
8. Report out as rare, few, moderate, or many. Be sure to report negative findings.

Note: If you are using a fixed-angle centrifuge, fill the tubes to within 1 cm of the top and centrifuge as above. Remove from centrifuge and place the tubes in a rack, fill to the top with additional flotation solution by gently running it down the inside edge to form a reverse meniscus, add coverslip and let stand for 5 or more minutes before reading.

For additional information and to order supplies contact:

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